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=> d his

     (FILE 'HOME' ENTERED AT 08:07:23 ON 06 NOV 2007)
     FILE 'REGISTRY' ENTERED AT 08:07:47 ON 06 NOV 2007
L1        STRUCTURE UPLOADED
L2      1 S L1
L3     49 S L1 FULL
          E ALEXA/CN
L4      1 S E15
L5      2 S E73-74
L6      1 S L3 AND L4
          SEL NAME L6
     FILE 'CA' ENTERED AT 08:16:44 ON 06 NOV 2007
L7    137 S L6 OR E1-2
L8     16 S ALEXA594 OR ALEXA FLUOR594 OR ALEXAFLUOR 594 OR ALEXAFLUOR594
L9    142 S L7-8
L10     5 S L8 AND PY<2003
L11    50 S L9 AND PATENT/DT
L12    25 S L11 AND PY<2005
     FILE 'BIOSIS' ENTERED AT 08:23:34 ON 06 NOV 2007
L13     5 S L10
     FILE 'MEDLINE' ENTERED AT 08:23:55 ON 06 NOV 2007
L14     5 S L10
     FILE 'CA, BIOSIS, MEDLINE' ENTERED AT 08:24:37 ON 06 NOV 2007
L15    33 DUP REM L10 L12 L13 L14 (7 DUPLICATES REMOVED)

=> d bib,ab,kwic l15 1-33

L15  ANSWER 19 OF 33  CA  COPYRIGHT 2007 ACS on STN
AN   138:283610  CA
TI   New fluorescent labeling technologies for ultrasensitive cytochemical
     and histochemical imaging
AU   Johnson, Iain
CS   Molecular Probes, Inc, USA
SO   Microscopy Today (2002), 10(4), 12, 14
AB   Mol. Probes, Inc. introduced two novel fluorescent labeling technologies
     for ultrasensitive cytochem. and histochem. imaging.  One of these is
     the Alexa Fluor series of dyes which optimizes spectroscopic and phys.
     properties that enable the prepn. of bioconjugates with consistently
     strong and photostable fluorescence output.  Functionally important
     characteristics of Alexa Fluor dyes include strong absorption at the
     output wavelengths of common excitation sources, resistance to
     photobleaching and self-quenching, and water soly. to facilitate
     coupling reactions with proteins and other biomols.  The well-
     differentiated spectra of the Alexa Fluor dyes provide many options for
     multicolor labeling and mol. proximity detection via fluorescence
     resonance energy transfer.  The second novel technol. is the Zenon
     immunolabeling technol. which provides a rapid, convenient and
     completely flexible technique for coupling dyes to antibodies.  Zenon
     technol. is based on dye- or enzyme-labeled Fab fragments of secondary
     antibodies directed against the Fc regions of primary antibodies.

L15  ANSWER 29 OF 33  CA  COPYRIGHT 2007 ACS on STN
AN   132:233984  CA
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TI Energy transfer compositions comprising phycobiliproteins
IN Haugland, Richard P.; Haugland, Rosaria P.
PA Molecular Probes, Inc., USA
SO PCT Int. Appl., 64 pp.

PI	WO 2000017650	A1	20000330	WO 1999-US22193	19990923
	WO 9915517	A1	19990401	WO 1998-US19921	19980923
PRAI	WO 1998-US19921	W	19980923		
	US 1998-209045	A	19981209		
	US 1997-935963	A	19970923		
	WO 1999-US22193	W	19990923		

AB Energy transfer compns. comprising one or more fluorescent dyes and a fluorescent protein are described, in particular where the fluorescent dye is a sulfonated dye and the fluorescent protein is a phycobiliprotein. The energy transfer compns. of the invention may further comprise addnl. fluorescent dyes or fluorescent proteins that act as intermediate energy transfer dyes or ultimate emitter dyes. The energy transfer compns. of the invention may also be substituted by chem. reactive functional groups, or covalently bound conjugated substances. The compns. of the invention possess utility as detection reagents and as fluorescent tracers in a wide variety of applications, including biol. applications.

L15 ANSWER 31 OF 33 CA COPYRIGHT 2007 ACS on STN
AN 134:97365 CA
TI Light-Induced Conformational Changes of Rhodopsin Probed by Fluorescent Alexa594 Immobilized on the Cytoplasmic Surface
AU Imamoto, Yasushi; Kataoka, Mikio; Tokunaga, Fumio; Palczewski, Krzysztof
CS Graduate School of Materials Science, Nara Institute of Science and Technology, Ikoma Nara, 630-0101, Japan
SO Biochemistry (2000), 39(49), 15225-15233
AB A novel fluorescence method has been developed for detecting the light-induced conformational changes of rhodopsin and for monitoring the interaction between photolyzed rhodopsin and G-protein or arrestin. Rhodopsin in native membranes was selectively modified with fluorescent Alexa594-maleimide at the Cys316 position, with a large excess of the reagent Cys140 that was also derivatized. Modification with Alexa594 allowed the monitoring of fluorescence changes at a red excitation light wavelength of 605 nm, thus avoiding significant rhodopsin bleaching. Upon absorption of a photon by rhodopsin, the fluorescence intensity increased as much as 20% at acidic pH with an apparent pKa of ~6.8 at 4°, and was sensitive to the presence of hydroxylamine. These findings indicated that the increase in fluorescence is specific for metarhodopsin II. In the presence of transducin, a significant increase in fluorescence was obsd. This increase of fluorescence emission intensity was reduced by addn. of GTP, in agreement with the fact that transducin enhances the formation of metarhodopsin II. Under conditions that favored the formation of a metarhodopsin II-Alexa594 complex, transducin slightly decreased the fluorescence. In the presence of arrestin, under conditions that favored the formation of metarhodopsin I or II, a phosphorylated, photolyzed rhodopsin-Alexa594 complex only slightly decreased the fluorescence intensity, suggesting that the cytoplasmic surface structure of metarhodopsin II is different in the complex with arrestin and transducin. These results demonstrate the

application of Alexa594-modified rhodopsin (Alexa594-rhodopsin) to continuously monitor the conformational changes in rhodopsin during light-induced transformations and its interactions with other proteins.

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STN INTERNATIONAL LOGOFF AT 08:25:24 ON 06 NOV 2007